

Original Article

Evaluating the Effectiveness of Silver Nanoparticle-Modified Dental Fillings on Enhancing Hard Tissue Durability

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ABSTRACT

This research investigates how the use of an etching gel containing silver nanoparticles (Ag NPs) affects the structural integrity and bonding strength at the “dentin-filling” and “enamel-filling” interfaces in permanent teeth with poor caries resistance in hard tissues. Teeth samples were taken from individuals aged 18 to 60 years, who underwent extractions for medical purposes. Adhesive systems from the IV and V generations were used for sealing, and both fluid and packable composites were selected as the filling materials. For metallographic analysis, the samples were cut perpendicular to the tooth axis, mechanically polished, and treated with concentrated orthophosphoric acid. Microscopic examinations were performed using both light and scanning electron microscopes. The findings show that the etching gel does not reduce the cohesive strength of the interfaces, even with the inclusion of silver nanoparticles.

Keywords: Etching gel, Cavity, Silver, Dental filling

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Introduction

The breakdown of the adhesive bond at the “dentin-filling” and “enamel-filling” junctions is a primary contributor to the formation of secondary caries, a problem that remains a significant challenge in dentistry [1-3]. A potential solution to mitigate this issue is remineralizing therapy, which helps maintain the bioorganic structure in dentin that has been affected by demineralization [4, 5]. Consequently, the addition of silver nanoparticles (Ag NPs) to dental materials

shows promise in combating secondary caries due to their potent antibacterial effects [6-8].

In healthcare, Ag NPs have a broad range of applications, including wound healing, surface sterilization, and coatings for implants [9-12]. The ongoing development of Ag NPs production methods is essential due to the imperfections of current technologies, with a focus on enhancing the stability and bioactivity of the products [13, 14]. However, the exact way in which Ag NPs affect the adhesive bonds in “dentin-filling” and “enamel-filling” remains

unclear, particularly whether silver-based substances weaken the cohesive strength of these boundaries. When applied, silver can penetrate the bioorganic matrix of demineralized dentin, contributing to its remineralization [15-18]. The etching process used in cavity preparation is key to ensuring the effective delivery of silver to both dentin and enamel [19-21]. This study aims to assess how an etching gel containing Ag NPs impacts the cohesive strength at the “dentin-filling” and “enamel-filling” interfaces in permanent teeth with low resistance to caries. The main focus of this research is to analyze the influence of an etching gel with silver nanoparticles on the strength properties of the “dentin-filling” and “enamel-filling” boundaries in permanent teeth from individuals with weakened caries resistance.

Materials and Methods

This study involved 46 teeth, including premolars and molars, extracted for medical reasons from patients aged between 40 and 60 years, all showing class I and II carious cavities [22]. The teeth were split into two groups, each containing 23 specimens. The first group, serving as the control, underwent the traditional treatment procedure, where the cavities were etched for fifteen seconds with a 36 percent H₃PO₄ solution [23]. In the second group, the teeth were treated for fifteen seconds with Etchmaster AgTM gel, which includes 36% H₃PO₄ and a filler with ten ppm of Ag NPs [12]. Both groups had their cavities filled with “AelitefloTM” fluid composite and “Aelite All-Purpose BodyTM” and “Aelite Aesthetic EnamelTM” packable composites, using adhesive systems of the IV generation “All-Bond 3TM” and V generation “Sealbond UltimaTM”. After the bonding process, both groups had their tooth and filling surfaces treated with the same gels used in each respective group, along with FortifyTM sealant. For metallographic analysis, the teeth were cut into 1 mm thick slices from the middle part of the crown, perpendicular to the tooth’s main axis (**Figure 1**). These slices were then polished mechanically, and the damaged surface was etched with concentrated H₃PO₄ for two to five minutes. The enamel near the filling interfaces and microstructure of the dentin was observed using a metallographic microscope at ×500 magnification. Higher magnification imaging was done using a scanning electron microscope (JSM-6390LV) [24]. To measure silver content in the teeth, an LSX-500 laser ablation unit (Cetac) was used [12].

Results and Discussion

The optical images captured at ×20 magnification revealed clear differences in color between the filling, enamel, and dentin, with their relative positions in the tooth crown reflecting the characteristics of the filling placement (**Figure 1**).

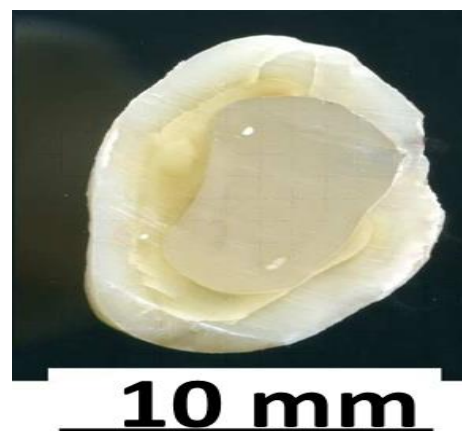
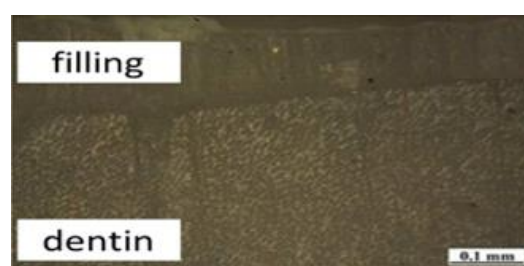
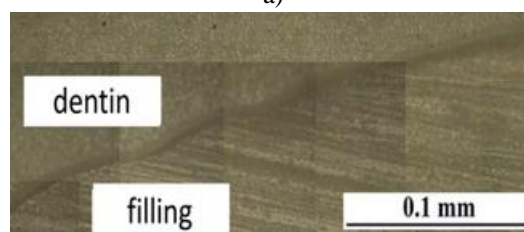


Figure 1. A sample cut from the crown of a tooth (×20)

The interfaces between the tooth’s hard tissues and the filling appeared as thin, uniform lines, free from pores or cracks. No noticeable differences were observed between the control groups. At higher magnification of ×500, observations confirmed that the “dentin-filling” (**Figure 2**) and “enamel-filling” interfaces remained intact, presenting smooth lines with no visible defects. A slight etching at the boundaries, which was undetectable at ×20 magnification, was attributed to the surface preparation method, which involved etching with concentrated orthophosphoric acid [25-28]. Similarly, no significant differences were found between the “dentin-filling” and “enamel-filling” boundaries in either group, whether in the control or the observation samples.



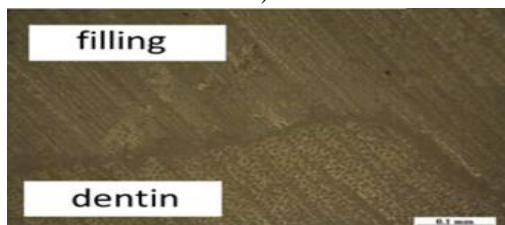
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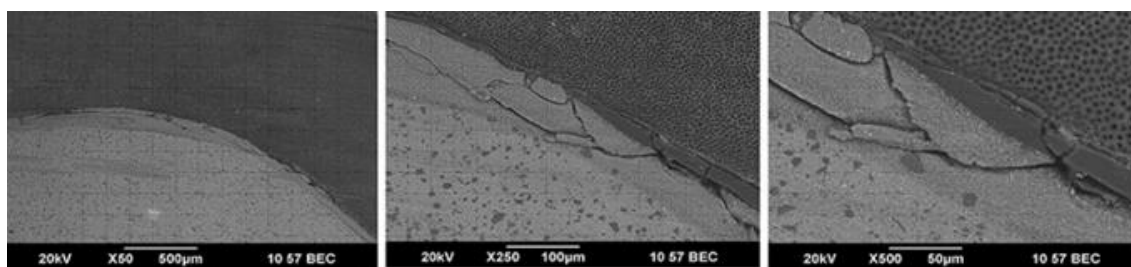
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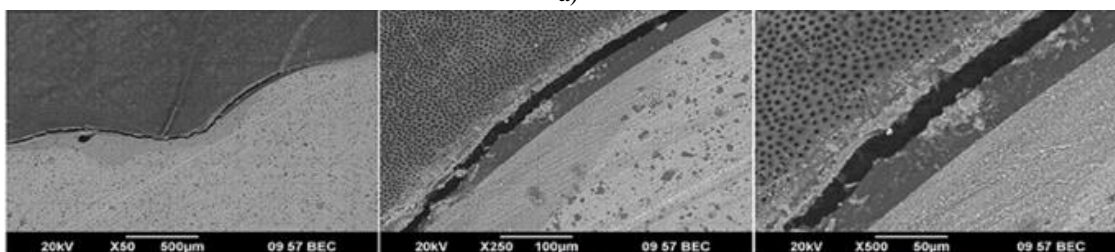
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Figure 2. The “dentin-filling” boundary after etching gel treatment (optical microscope); a) With Ag NPs, using an adhesive system from the IV generation, b) With Ag NPs, using an adhesive system from the V generation, c) Without Ag NPs, using an adhesive system from the IV generation, d) Without Ag NPs, using an adhesive system from the V generation.

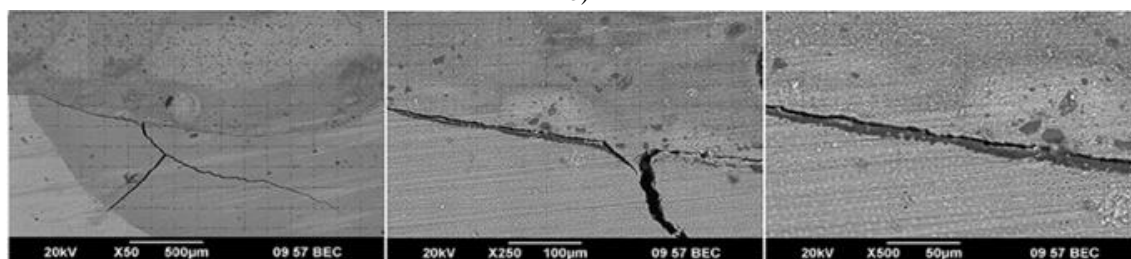
The findings from the electron microscopy align with the previously mentioned observations. The interfaces of the “dentin-filling” (**Figure 3**) and “enamel-filling” (**Figure 4**) were free from continuity defects, showing uniform etching across their entire length. The presence of a few microcracks at the boundaries is likely due to mechanical stress applied during the tooth-cutting process and the mechanical polishing of the sample surface [29-31]. Additionally, factors like acid etching and dehydration during storage could contribute to the formation of these cracks, possibly leading to sample deformation and weakening of the adhesive bond [32-35]. However, it is important to note that these cracks do not extend into the surface layer of the dentin or enamel. Despite enduring substantial mechanical forces, none of the samples from the structural studies showed failure at the boundaries [36-38]. Notably, electron microscopy revealed no significant differences in the boundaries between samples from the observation and control group.



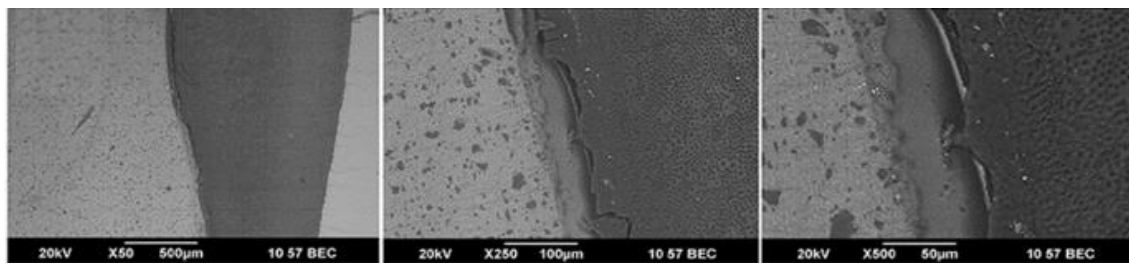
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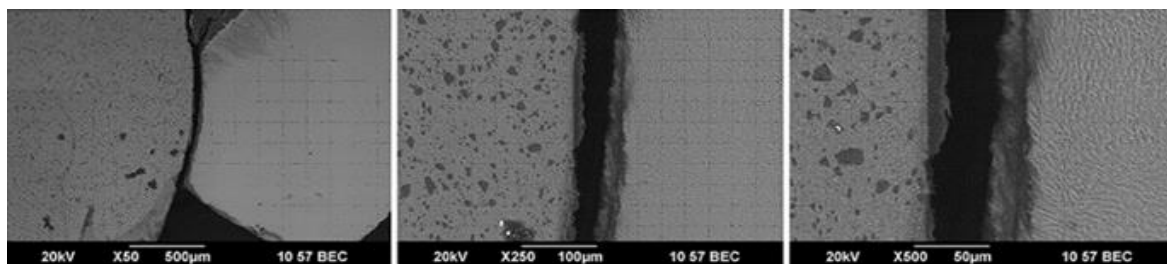


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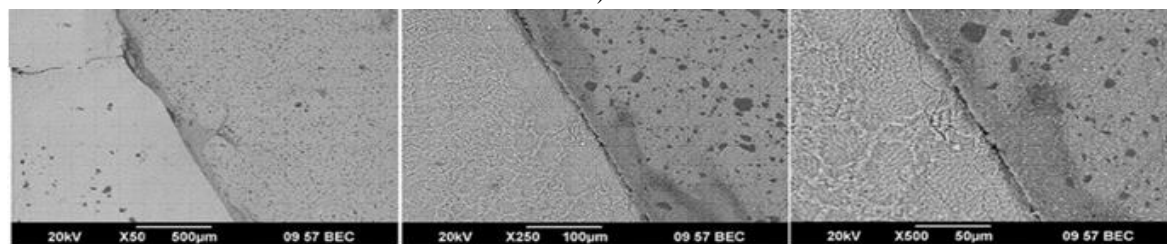


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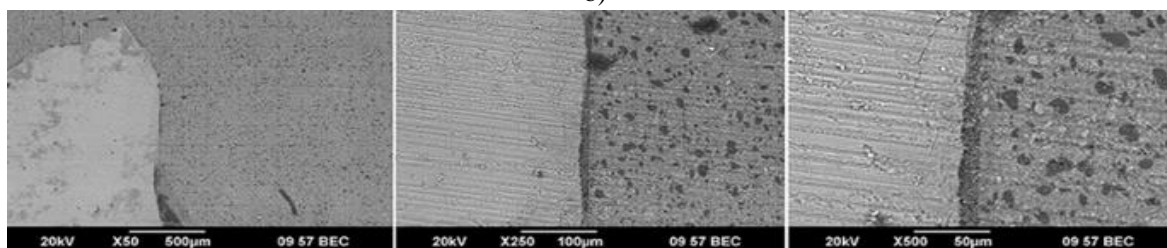
Figure 3. The “dentin-filling” boundary following etching gel treatment (scanning electron microscope); a) With Ag NPs, using an IV generation adhesive system, b) With Ag NPs, using a V generation adhesive system, c) Without Ag NPs, using an IV generation adhesive system, and d) Without Ag NPs, using a V generation adhesive system.



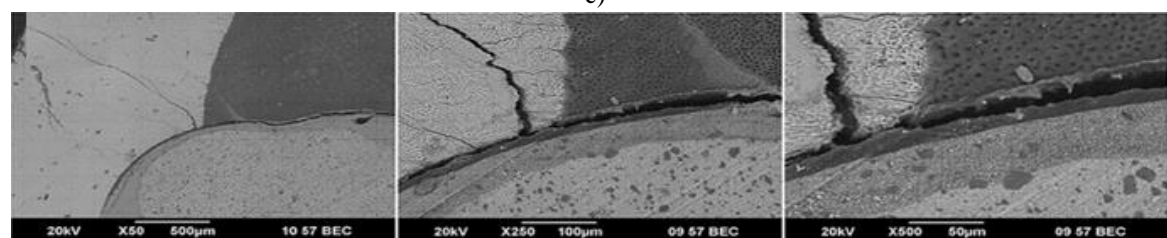
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Figure 4. The “enamel-filling” boundary after etching gel treatment (scanning electron microscope)*; a) With Ag NPs, using an IV generation adhesive system, b) With Ag NPs, using a V generation adhesive system, c) Without Ag NPs, using an IV generation adhesive system, and d) Without Ag NPs, using a V generation adhesive system.

The analysis of the elemental composition of dentin and enamel in the control group samples performed approximately 1.25 mm away from the filling

interface, revealed that a 50 µm thick layer of hard tissue contained around 10 ppm of silver (**Figure 5**). In

contrast, no silver was detected in the samples from the observation group.

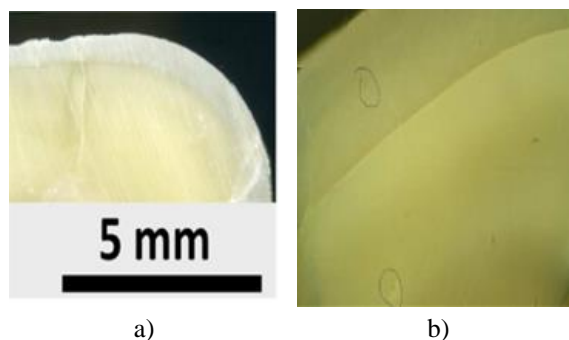


Figure 5. Dentin-enamel junction treated with etching gel containing Ag NPs; a) After cutting (showing no cracks or pores at the boundary), and b) After analyzing the elemental composition in dentin and enamel (laser ablation sites are marked).

The examination of the cohesive strength at the “dentin-filling” and “enamel-filling” interfaces using physical materials science techniques provided two important insights:

The cohesive strength at these boundaries was strong enough to resist the mechanical forces applied during sample cutting and polishing. Also the impact of aggressive treatments like concentrated orthophosphoric acid, regardless of the etching gel contained Ag NPs.

When the etching gel containing Ag NPs was used, a layer of hard tissue, located about 1.25 millimeters from the boundaries of the prepared cavities, became enriched with Ag NPs. These levels of silver were found to be sufficient to produce bactericidal and fungicidal effects [39, 40].

Conclusion

The results of this study indicate that the presence of up to 10 ppm of Ag NPs in enamel and dentin does not impact the restoration’s strength. As such, the primary role of silver in the etching gel appears to be its preventive effect on pathology.

However, it is essential to remember that, in clinical practice, the bond strength between the filling and the tooth’s hard tissues can be influenced by various factors. These include the condition of the enamel and dentin, the specific properties of the filling material, and the chemical composition of both the etching gel and the adhesive system. The study used samples from teeth extracted for medical purposes, which closely mimics natural conditions. Given the results, it can be noticed that Ag NPs in the etching gel serve mainly as

a preventive agent against pathology without affecting the restoration’s strength properties.

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Conflict of Interest: None

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Ethics Statement: None

References

1. Immich F, de Araújo LP, da Gama RR, da Rosa WLO, Piva E, Rossi-Fedele G. Fifteen years of engine-driven nickel-titanium reciprocating instruments, what do we know so far? An umbrella review. *Aust Endod J.* 2024;50(2):409-63. doi:10.1111/aej.12870
2. Milian R, Lefrançois E, Radzikowski A, Morice S, Desclos-Theveniau M. Pre-orthodontic restorative treatment of microdontia diastema teeth using composite injection technique with a digital workflow-case report. *Heliyon.* 2023;9(5):e15843. doi:10.1016/j.heliyon.2023.e15843
3. Awad R, Musa M, Elhoumed M, Liu F, Guo Q. Comparison of endoflas and zinc oxide eugenol as root canal filling materials for pulpectomy in deciduous teeth: a systematic review and meta-analysis. *Saudi Dent J.* 2024;36(6):821-9. doi:10.1016/j.sdentj.2024.03.007
4. Khurshid Z, Adanir N, Ratnayake J, Dias G, Cooper PR. Demineralized dentin matrix for bone regeneration in dentistry: a critical update. *Saudi Dent J.* 2024;36(3):443-50. doi:10.1016/j.sdentj.2023.11.028
5. Barbosa CB, Monici Silva I, Dame-Teixeira N. The action of microbial collagenases in dentinal matrix degradation in root caries and potential strategies for its management: a comprehensive state-of-the-art review. *J Appl Oral Sci.* 2024;32(8):e20240013. doi:10.1590/1678-7757-2024-0013
6. Elmarsafy SM. A comprehensive narrative review of nanomaterial applications in restorative dentistry: demineralization inhibition and remineralization applications (Part I). *Cureus.* 2024;16(4):e58544. doi:10.7759/cureus.58544
7. Wang K, Wang S, Yin J, Yang Q, Yu Y, Chen L. Long-term application of silver nanoparticles in dental restoration materials: potential toxic injury to the CNS. *J Mater Sci Mater Med.* 2023;34(11):52. doi:10.1007/s10856-023-06753-z

8. Bolenwar A, Reche A, Dhamdhare N, Rathi S. Applications of silver nanoparticles in dentistry. *Cureus*. 2023;15(8):e44090. doi:10.7759/cureus.44090
9. Vanlalveni C, Ralte V, Zohmingliana H, Das S, Anal JMH, Lallianrawna S, et al. A review of microbes mediated biosynthesis of silver nanoparticles and their enhanced antimicrobial activities. *Heliyon*. 2024;10(11):e32333. doi:10.1016/j.heliyon.2024.e32333
10. Akhter MS, Rahman MA, Ripon RK, Mubarak M, Akter M, Mahbub S, et al. A systematic review on green synthesis of silver nanoparticles using plants extract and their bio-medical applications. *Heliyon*. 2024;10(11):e29766. doi:10.1016/j.heliyon.2024.e29766
11. Kshatriya VV, Kumbhare MR, Jadhav SV, Thorat PJ, Bhambarge RG. An updated review on emerging recent advances and biomedical application of silver nanocluster. *Zhongguo Ying Yong Sheng Li Xue Za Zhi*. 2023;39:e20230001. doi:10.62958/j.cjap.2023.001
12. Blinov AV, Nagdalian AA, Povetkin SN, Gvozdenko AA, Verevkina MN, Rzhepakovsky IV, et al. Surface-oxidized polymer-stabilized silver nanoparticles as a covering component of suture materials. *Micromachines (Basel)*. 2022;13(7):1105. doi:10.3390/mi13071105
13. Ahmadi M, Sabzini M, Rastgordani S, Farazin A. Optimizing wound healing: examining the influence of biopolymers through a comprehensive review of Nanohydrogel-embedded nanoparticles in advancing regenerative medicine. *Int J Low Extrem Wounds*. 2024;15347346241244890. doi:10.1177/15347346241244890
14. Mustafa S, Alharbi LM, Abdelraheem MZ, Mobashar M, Qamar W, A Al-Doaiss A, et al. Role of silver nanoparticles for the control of anthelmintic resistance in small and large ruminants. *Biol Trace Elem Res*. 2024;202(12):5502-21. doi:10.1007/s12011-024-04132-5
15. Nizami MZI, Xu VW, Yin IX, Yu OY, Chu CH. Metal and metal oxide nanoparticles in caries prevention: a review. *Nanomaterials (Basel)*. 2021;11(12):3446. doi:10.3390/nano11123446
16. Yin IX, Zhao IS, Mei ML, Li Q, Yu OY, Chu CH. Use of silver nanomaterials for caries prevention: a concise review. *Int J Nanomed*. 2020;15:3181-91. doi:10.2147/IJN.S253833
17. Afkhami F, Forghan P, Gutmann JL, Kishen A. Silver nanoparticles and their therapeutic applications in endodontics: a narrative review. *Pharmaceutics*. 2023;15(3):715. doi:10.3390/pharmaceutics15030715
18. Xu GY, Zhao IS, Lung CYK, Yin IX, Lo ECM, Chu CH. Silver compounds for caries management. *Int Dent J*. 2024;74(2):179-86. doi:10.1016/j.identj.2023.10.013
19. Asghar M, Omar RA, Yahya R, Yap AU, Shaikh MS. Approaches to minimize tooth staining associated with silver diamine fluoride: a systematic review. *J Esthet Restor Dent*. 2023;35(2):322-32. doi:10.1111/jerd.13013
20. Burgess JO, Vaghela PM. Silver diamine fluoride: a successful anticariogenic solution with limits. *Adv Dent Res*. 2018;29(1):131-4. doi:10.1177/0022034517740123
21. Vornic I, Pop O, Pascalau A, Andreescu G, Beiusan C, Manole F, et al. Assessing the role of Bcl-2 and p53 in apoptotic mechanisms in spontaneous abortions. *Pharmacophore*. 2024;15(2):1-6. doi:10.51847/CO2qttSgIN
22. Mount GJ, Tyas JM, Duke ES, Hume WR, Lasfargues JJ, Kaleka R. A proposal for a new classification of lesions of exposed tooth surfaces. *Int Dent J*. 2006;56(2):82-91. doi:10.1111/j.1875-595x.2006.tb00078.x
23. Patel SR, Jarad F, Moawad E, Boland A, Greenhalgh J, Liu M, et al. The tooth survival of non-surgical root-filled posterior teeth and the associated prognostic tooth-related factors: a systematic review and meta-analysis. *Int Endod J*. 2024;57(10):1404-21. doi:10.1111/iej.14116
24. Kravtsov AA, Blinov AV, Nagdalian AA, Gvozdenko AA, Golik AB, Pirogov MA, et al. Acid-base and photocatalytic properties of the CeO₂-Ag nanocomposites. *Micromachines (Basel)*. 2023;14(3):694. doi:10.3390/mi14030694
25. Al-Khateeb SN, Tarazi SJ, Al Maaitah EF, Al-Batayneh OB, Abu Alhaija ES. Does acid etching enhance remineralisation of arrested white spot lesions? *Eur Arch Paediatr Dent*. 2014;15(6):413-9. doi:10.1007/s40368-014-0131-2
26. Bataineh M, Malinowski M, Duggal MS, Tahmassebi JF. Comparison of the newer preventive therapies on remineralisation of enamel in vitro. *J Dent*. 2017;66:37-44. doi:10.1016/j.jdent.2017.08.013
27. Kallagova AR, Bulgakova YV, Oganyan IG, Mussakaeva KH, Adoneva VA, Burlakova TO. Effectiveness of fluoridated milk in children's daily diet for the prevention of caries. *Arch Pharm*

- Pract. 2023;14(3):26-32. doi:10.51847/0BB2ttxz1A
28. Green S, Carusi A, Hoeyer K. Examining the role and efficiency of personalized medicine in the diagnosis, prevention, and treatment of diseases. *Int J Pharm Res Allied Sci.* 2024;13(2):104-13. doi:10.51847/OibXYhUnry
 29. Sedani S, Kriplani S, Thakare A, Patel A. The hidden world within: microbial dynamics in root canal systems. *Cureus.* 2024;16(5):e60577. doi:10.7759/cureus.60577
 30. Nikkerdar N, Golshah A, Salmani Mobarakeh M, Fallahnia N, Azizi B, Shoochanizad E, et al. Recent progress in application of zirconium oxide in dentistry. *J Med Pharm Chem Res.* 2024;6(8):1042-71. doi:10.48309/jmpcr.2024.432254.1069
 31. Khyade VB, Yamanaka S, Bajolge R. Utilization of BSF-cream for antiaging impact on human skin. *Entomol Appl Sci Lett.* 2024;11(1):56-66. doi:10.51847/BdrxgiFL1L
 32. Sampoerno G, Sukaton S, Ferdinandus E, Firdaus N, Damayanti R. Expression of CGRP and NaV 1.8 in neurons and macrophages after p.gingivalis lipopolysaccharide application on dental pulp tissue. *J Med Pharm Chem Res.* 2024;6(5):558-70. doi:10.48309/jmpcr.2024.187568
 33. Mokeem LS, Garcia IM, Melo MA. Degradation and failure phenomena at the dentin bonding interface. *Biomedicines.* 2023;11(5):1256. doi:10.3390/biomedicines11051256
 34. Fang K, Chen K, Shi M, Wang L. Effect of different adhesive systems on dental defects and sensitivity to teeth in composite resin restoration: a systematic review and meta-analysis. *Clin Oral Investig.* 2023;27(6):2495-511. doi:10.1007/s00784-023-05007-0
 35. Nurmuhambetov IR, Prokopovich DS, Chernishev AE, Shorganova AA, Samarkin SV, Magomadova TT, et al. Assessment of the effect of biocompatibility of fibroblasts and scaffolds on the cell cycle in vitro. *J Adv Pharm Educ Res.* 2024;14(3):49-53. doi:10.51847/tbAag0pml3
 36. Ng TC, Chu CH, Yu OY. A concise review of dental sealants in caries management. *Front Oral Health.* 2023;4:1180405. doi:10.3389/froh.2023.1180405
 37. Cadenaro M, Josic U, Maravić T, Mazzitelli C, Marchesi G, Mancuso E, et al. Progress in dental adhesive materials. *J Dent Res.* 2023;102(3):254-62. doi:10.1177/00220345221145673
 38. Nica-Badea D. Impact of translocation and bioconcentration of heavy metals in the area of lignite-fired power plant. *J Biochem Technol.* 2024;15(2):51-8. doi:10.51847/otraDHXihN
 39. Gudkov SV, Serov DA, Astashev ME, Semenova AA, Lisitsyn AB. Ag2O nanoparticles as a candidate for antimicrobial compounds of the new generation. *Pharmaceuticals (Basel).* 2022;15(8):968. doi:10.3390/ph15080968
 40. Al-Twajiri SA, AlKharboush GH, Alohal MA, Arab IF, Alqarni RH, Alharbi MS. Application of lasers for soft tissues in orthodontic treatment: a narrative review. *Bull Pioneer Res Med Clin Sci.* 2024;3(1):1-6.